

6.4 kb

Book N. _____

1/6/95

155

age No. _____

Purpose: To repeat & optimize previously 6.4 kb.

Tried miniprep DNA as a control

Will try 2 dif. cycling conditions 3 step as well as 2 step.

Colonies will be lysed in 2 different ways 1. in PK (single
2. in H₂O colony buffer

Unlysed will also be included again.

Conditions - since two PCR dNTP + 2 mM Mg²⁺, pH 7.5, 10 mM Tris-HCl
+ 1 mM EDTA
worked with Tag 5'V, the same 50 µg/ml PK

Centrifugation will be used.

Need 2 µl g miniprep - can unknown (need BM3 ③)
still have to run agar

Tried dif. enzyme conc 1:2, 5 and 1:10, 2:10, 5:10

Tag

Tag 5' V

Colonies lysis: Since these colonies were so small after 37° pooled 5 or 6 colonies in a single area -
spotted 2 µl g lysis buffer or H₂O mixed & pipetted out the liquid on to a tube containing 13 µl g lysis buffer
or H₂O

Colonies in PK lysis 55°, 15', → 95°, 15'

in H₂O : 95°, 15'

(Added a 5 µl g H₂O) pooled all three tubes together
and made up the volume to 50 µl.

Should have picked more for more reactions

used 10 µl / Pix - apparently either PK lysed or H₂O lysed &
very slowly from 14 to 16 hrs To Page No. _____

Ised & Understood by me,

Date

Invent'd by

Date

1/6/95

Recorded by

1/6/95

K. Subramaniam

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For mini-prep DNA : prepare premix with template

For colony : add them later

miniprep: premix : 25 x

A

dNTP 25 μ l (200 μ M each / Rx)

F.P 5 (2.0 μ M) 0.4 μ M

R.P 5 0.4 μ M

Template 25 \times 2 \rightarrow last expt used 1.5 + 1 / 2 x

miniprep = 2 μ l / Rx.

 H_2O

415

500 \rightarrow 25 μ l / Rx

Premix B: 5 x

Tag

5

Tag + DN

1 : .01 2 : .02 5 : .05

(2 mM) Buffer B

50

100 \times enzyme

1 2.5

5

10

25

 H_2O

99.5

99

97.5

95

90

75

150

\downarrow 30 μ l / Rx.

3 step
2 cycle

2 step:

94° 3'

94° 3'

20

20 (94° 45'
55° 30'
72° 3')

94° 45'

68° 5'

T Pag 1

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Date

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Dat

19/05

R c rded by

R. Sureshwaran

1/6/94

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2 step

Tube #	2 step
1	1
2	1
3	2
4	2
5	5
6	5
7	1 : 01
8	1 : .01
9	2 : .02
10	2 : .02
11	5 : .05
12	5 : .05

3 step

Tube #	3 step
	13
	14
	15
	16
	17
	18
	19
	20
	21
	22
	23
	24

Same as in 2 step.

columns1 mix A: 15 xMix B: 5 x as earlier
for T + S.V

dNTP 1.5

primers 3

Taq alone and left over
from previous
page

" 3

- 150 (Template 10 μl / rx when added later)

120 1.29

4

- 150 → 10 μl / rx

20 μl

→ + ←

added 30 μl / rx

approximately either

Taq alone or Taq + DV

changing condition same as on previous page.

for 3 step cycle, 2 step not done. ∵ didn't
have much template left from

mix 25 1 + .01 }

PK lysed

lysed plasmid

26 2 + .02 }

31)

27 5 + .05 }

28) 11/20

32)

35, 36 (20 Taq)

29) lysed

33)

straight pick

How plain

30)

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Ised & Understood by me,

Date

Invented by

Date

11/16/95

Recorded by

11/16/94

K. Sitaraman

Project No. _____

158 (444)

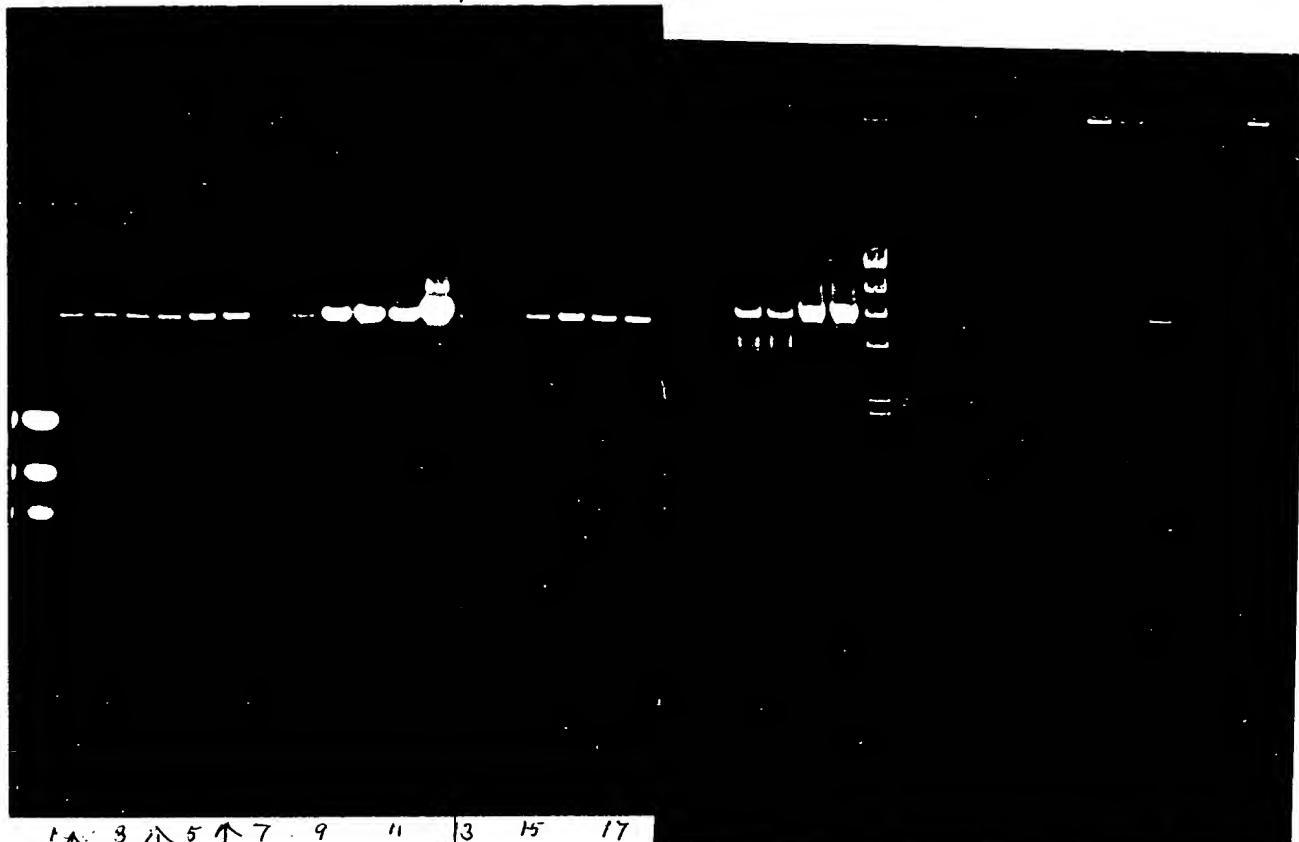
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Step
2 cycle

3 step cycle



1 ↑ 3 ↑ 5 ↑ 7 ↑ 9 ↑ 11 ↑ 13 ↑ 15 ↑ 17 ↑

Tag

19 ↑ 29 ↑ 23 ↑
T + D ✓

pk direct
 H_2O peak

1
2
3 Enzyme mix

Unit 1st
enzymatic
Tag

5 1, 2, 5

pk direct
 H_2O peak w/ Tag alt
typed typed)
w/ Tag + DV

mini prep.

plasmids

Result: Even 3 step gave better product with less noise.
plasmid amp should be done under more control.

T Pag 1

Witnessed & Understood by me,

Date

1/20/95

Invented by

Rec'd by

K. Stareman

Date

1/9/95